ORIGINAL ARTICLE

Up-Regulation of Interleukins (ILS) and their Potential Role in the Pathogenesis of Radicular Cyst

ANOOSH QAYYUM¹, AMINA SHAHID², KHALIDA ANWAR³, NUSRAT TARIQ⁴, SULTAN AHMAD⁵, NAVEED SHUJA⁶, SARA ZAHID⁷, SULAYMAN WAQUAR⁷, ARIF MALIK ⁷

¹Assistant Professor Department of Biochemistry, Rawalpindi Medical University, Rawalpindi-Pakistan

²Assistant Professor Department of Biochemistry, Pak Red Crescent Medical College Dina Nath, Lahore-Pakistan

³Assistant Professor Department of Biochemistry, RYK Medical College and Allied hospitals, Rahim Yar Khan-Pakistan

⁴Professor of Physiology Department, M.Islam Medical and Dental College, Gujranwala-Pakistan

⁵Associate Professor Department of Physiology, Karachi Institute of Medical sciences, Karachi-Pakistan

⁶Associate Professor Department of Biochemistry, Lahore Medical and Dental College, Lahore-Pakistan

⁷Institute of Molecular Biology and Biotechnology (IMBB), The University of Lahore-Pakistan Corresponding author: Arif Malik, Email: arifuaf@yahoo.com, Cell: + 92321-8448196

ABSTRACT

Objective: Study the upregulation of Interleukin levels and their potential role in the pathogenesis of radicular cysts **Study Design:** Cross-sectional study

Place and Duration: Institute of Molecular Biology and Biotechnology (IMBB), The University of Lahore, (9-10 Months)

Martials and Methods: Hundred samples were extracted from both diseased and healthy individuals and tissue homogenates were prepared. MDA and LPS were estimated with the help of spectrophotometric method. IL1, IL-6, TNF-α and MMP-9 were determined by their commercial ELISA kits.

Results: Malondialdehyde (MDA) was significantly (p=0.033) higher (3.99 ± 0.99 Vs .95±0.011) in diseased group as compared to control. Higher levels of IL-1 (pg/ml) (6.59 ± 1.56 Vs 4.56 ± 1.23), IL-6 (pg/ml) (7.65 ± 1.41 Vs 5.99 ± 0.219) and TNF- α (pg/ml) (31.55 ± 4.55 Vs 20.55 ± 3.28) were recorded in the diseased group. In the group of patients, MMP-9 remained significantly increased when compared with the healthy controls in the case of subjects it remained (93.25 ± 6.48 ng/ml) while in controls they were (47.22 ± 5.29 ng/ml). Likewise in the case of LPS it was significantly increased in the group of radicular cyst (17.25 ± 4.26 pg/ml) as compared to healthy controls (125.25 ± 9.28 pg/ml).

Conclusion: Present study indicates the significant correlation of LPS with the pathogenesis of radicular cyst. Moreover, it indicates that as the oxidative stress is increased, it increases the expression of several ILs and MMP-9 that contribute in the pathogenesis of radicular cyst. Further studies may describe the role of lipopolysaccharide and other potential variables to enable better understanding about bone resorption mechanism in periapical cyst development.

Keywords: Interleukins, Lipopolysaccharides (LPS), Malondialdehyde (MDA), MMP-9, Radicular Cyst.

INTRODUCTION

The tooth associated with chronic periapical lesion shows the long lasting process of infection due to a number of microorganisms existing in root canal system and depends on host response¹. In oral disease, anaerobic gram negative bacteria and endotoxin play а vital role to cause chronic apical periodontitis². Lipopolysaccharide (LPS) released from gram negative bacteria mediates the process of disease around the root apex. In periapical lesion, LPS shows a positive correlation with disease cruelty³. Apical periodontitis is a chronic disease initiated by the microbial action on pup tissue causing its death and the movement of bacteria towards the apical periodontal tissue that attaches teeth to the alveolar bone. This morbid condition characterizes local inflammatory response due to the presence of large amount of microorganisms and their distribution from infected root canal towards apical and periapical tissue resulting in bone resorption in periradicular area^{4,24}.

Microorganisms i.e. gram negative bacteria instead of having severe harmful features and yielding toxic products and biproducts for apical and periapical tissue comprise endotoxin in their outer wall⁵. Bacterial proliferation and death accelerates the release of large amount of endotoxin from their wall which ultimately leads to the inflammation and resorption of bone. Endotoxin comprises of lipids, proteins and polysaccharides and thought to be one of the most effective microbial weapon playing a key role in the development of radicular cyst^{6,23}. LPS has no direct effect on tissue damage, it stimulates different cells such as macrophages, neutrophils, fibroblasts, T and B lymphocytes and plasmocytes. These inflammatory cells release various cytokines including interferon, TNF- α , interleukins, different growth factors and prostaglandins⁷ which may take part in local tissue injury. Local production of bone resorbing cells and their stimulus are obligatory for alveolar bone loss around the tooth apex. Bone resorption is mediated by different factors such as interleukins (IL-1, IL-6, IL-11 and IL-17), TNF- α, TNF-β, nitric oxide, endothelin,

prostaglandins, lipopolysaccharides, reactive oxygen species (ROS) and matrix metalloproteinases^{8,22}.

In periapical disease, resorption of bone is caused by the cooperation of both innate and adaptive immune response. Polymorphonuclear leukocytes (PMNs), macrophages and lymphocytes are formed in apical periodontal tissue during chronic inflammation and these cells are potent inducer of bone resorbing cytokines (IL-1, IL-6 and TNF- α) stimulating the bone resorption⁹. Activation and generation of osteoclasts influenced by TNF family of cytokines are produced by lymphocytes of adaptive immune response. Evidence suggests that the activation of lymphocytes by the stimulus of antigen in adaptive immune response play a major role in periapical bone loss¹⁰. Matrix metalloproteinases (MMPs) are zinc dependent endopeptidases that can degrade all types of extracellular matrix. Collagenases (MMP-1, MMP-8 and MMP-13) are important to break the fibrillar collagen and these small fragments of collagens are further degraded by gelatinases (MMP-2 and MMP-9). These MMPs are involved in the destruction of different periapical lesions and peri-implantitis disease^{26, 27}. The aim of current study was to investigate the role of lipopolysaccharide, matrix metalloproteinase-9 and inflammatory cytokines in periapical bone resorption during the development of radicular cyst.

MATERIALS AND METHODS

Hundred (n=100) male tissue homogenates of radicular cyst associated with root canal treated teeth (RCT) and hundred (n=100) normal healthy pulp tissue samples were taken from extracted teeth for orthodontic treatment from the de'Montmorency College of Dentistry Lahore. Inclusion and exclusion criteria of the study remain as the patients having periodontal issues especially radicular cyst were included in the following study while those having any other congenital diseases such as thyroids defects and diabetes were excluded out of the study, all of the work was approved by Research and Ethics Committee at University of Lahore. Tissue homogenates were collected in appropriate containers and stored at required temperature for their future assay. Levels of ILs, TNF- α , MMP-9 were estimated by their ELISA whereas, MDA and LPS were estimated with spectrophotometric methods respectively³⁸.

Mean weight (kg), age (years), blood pressure (mmHg), BMI (kg/m²), complete blood count of the controls and patients were estimated that show statistically insignificant change in weight, age, BMI, blood pressure whereas, intensity of Hb, Hct and neutrophils were significantly different in controls and patients.

Statistical Analysis: Statistical analysis was done by using SPSS (v.16), applied tests were Independent t-test and Pearson correlation. Results were expressed as (Mean \pm S.D) taking (p<0.05) as significant.

RESULTS

Results of table 1 shows the demographic distribution of the variables among the radicular cyst patients. Table shows significant differences among the variables in the patients with radicular cysts when were compared with the healthy individuals. The level (nmol/ml) of Malondialdehyde (MDA) was significantly (p=0.033) higher (3.99±0.99 Vs .95±0.011) in diseased group as compared to control. The role of inflammatory cytokines was recognized as medically potential analytical variable and used to establish the degree of disease severity, stage and treatment protocols. The data presented in table regarding tested inflammatory cytokines (IL-1 and TNF- α) depicted that these cytokines play a very important role in the development of a radicular cyst. Higher levels of IL-1 (pg/ml) (6.59±1.56 Vs 4.56±1.23), IL-6 (pg/ml) (7.65±1.41 Vs 5.99±0.219) and TNF-α (pg/ml) (31.55±4.55 Vs 20.55±3.28) were recorded in the diseased group as compared to normal and differed significantly from each other. In the group of patients, MMP-9 remained significantly increased when compared with the healthy controls in the case of subjects it remained (93.25±6.48 ng/ml) while in controls they were (47.22±5.29 ng/ml). Likewise, in the case of LPS it was significantly increased in the group of radicular cyst (17.25±4.26 pg/ml) as compared to healthy controls (125.25±9.28 pg/ml) as shown in table 2.

Table 1: Demographic and Hematological Profile in Patients with Radicular Cyst

Variables	Control (n=100)	Subjects (n=100)	P-value (<0.05)
WEIGHT (Kg)	71.33±3.26	70.29±6.26	0.589
AGE (Years)	47.21±4.55	48.29±4.29	0.581
SBP (mmHg)	120.22±1.99	123.26±1.88	0.226
DBP (mmHg)	80.26±4.26	81.26±2.26	0.4.26
BMI (kg/m ²)	21.26±1.56	22.26±4.26	0.421
RBC (M/mcl)	4.16±0.98	4.55±0.66	0.716
WBC (k/mcl)	7.16±0.62	8.16±1.14	0.039
Hb (g/dl)	13.26±1.55	12.56±2.61	0.253
PLT (k/mcl)	299.23±4.26	289.23±4.26	0.532
Hct (%)	39.26±2.66	46.23±4.55	0.032
NEUTROPHILS (%)	60.99±1.99	102.36±2.88	0.016

Table 2: Levels of Different Variables in Radicular Cyst

Variables	Control (n=100)	Subject (n=100)	P- value
MDA (nmol/ml)	0.95±0.011	3.99±0.99	0.004
IL-1 (pg/ml)	4.56±1.23	6.59±1.56	0.019
IL-6 (pg/ml)	5.99±0.21	7.65±1.41	0.000
TNF-α (pg/ml)	20.55±3.28	31.55±4.55	0.011
MMP-9 (ng/ml)	47.22±5.29	93.25±6.48	0.000
Lipopolysaccharides (pg/ml)	17.25±4.26	125.25±9.28	0.028

DISCUSSION

In response to gram negative bacterial infection, a large amount of LPS is produced in the pulp canal and around the tooth apex. The existence of LPS may be a prerequisite for the development and progression of periapical lesion and bone resorption because it

excites the inflammatory cells to release different proinflammatory cytokines which cause local tissue injury¹¹. The magnitude of tissue injury originated by LPS depends on host innate immune response. In periapical area, inflammatory reaction encompasses the recruitment of neutrophils, monocytes, macrophages, lymphocytes, fibroblasts and plasma cells to secrete interleukins (IL-1 and IL-6), TNF-α, iNOS, MMP-9 and ROS^{12,19}. After bacterial infection, the initial step of immune response is to recognize the bacterial pathogen i.e. LPS by using pathogen associated molecular patterns (PAMPs). The host having a protein called lipoprotein binding protein (LBP) produced by hepatocytes in the liver during chronic inflammation, binds to LPS and facilitates its transfer towards CD14 receptor^{13,21}. CD14 receptors are expressed by macrophages, neutrophils, epithelial cells, fibroblasts and endothelial cells^{14,20}. Studies show that the definite receptor of LPS is toll like receptor-4 (TLR4) and CD14 binding with LPS presents it to human myeloid differentiating protein (MD2) and ultimately to TLR415,25

LBP binding with LPS activates CD14/MD2/TLR4 complex which is critical for sensitivity and activation of intracellular signalling pathways to stimulate the production of inflammatory cytokines and chemical mediators¹⁶. Previous studies show that the activation of nuclear factor kappa B (NFκβ) is crucial for the increased expression of pro-inflammatory cytokines, receptor activator of nuclear factor kappa-B ligand (RANKL) and matrix metalloproteinases. The increased production of proinflammatory cytokines, RANKL and MMPs by LPS is crucial for degradation of the bone and periradicular tissue during cyst expansion^{17,18}. Martinho et al. showed that when the size of periapical lesion was compared with the production of IL-1 and IL-6, it was concluded that large sized periapical lesions had increased production of these cytokines²⁸. Current study suggests that these cytokines are associated with increased tissue injury at the site of inflammation. Furthermore, IL-1 and IL-6 may directly cause the activation and production of osteoclasts by stimulating the transcription factor NF κ B²⁹. Wan et al. concluded that TNF- α , IL-1 and IL-6 may stimulate the differentiation of osteoclasts in a synergic way and they may interact among each other to enhance the periapical bone resorption^{30, 31}.

TNF- α and IL-1 are found to stimulate the increased expression of RANKL on the surface of osteoblasts which is a key factor for competitive binding to RANK on the surface of osteoclast replacing osteoprotegerin (TNF-α vs RANKL, r= 0.644** and IL-1 vs RANKL, r=0.765**) . This binding of RANKL which is a ligand for its receptor RANK initiates the activation of preosteoclasts to mature osteoclasts ³². These activated osteoclasts attach with bone and release acid which demineralize the inorganic part of bone and organic matrix of bone is degraded by matrix metalloproteinases³³. The oral bacteria present in infected root canal triggers the production of MMPs by macrophages and neutrophils in the periapical area. The aim of root canal treatment is to eliminate bacteria, toxins, virulent factors and chronic inflammation in the peri-radicular area. As the root canal is free from bacteria and its toxins the amount of cytokines, MMPs and inflammatory cells in the periradicular area is also decreased^{34,35}. Hence, IL-1, IL-6 and TNF- α dependent periapical bone loss and MMP-9 dependent tissue injury is also decreased. IL-1 secreted by epithelial cells of periapical cyst and inflammatory cells not only upregulates the pro MMP-9 secretion but also activates MMP-9 by autocrine and paracrine signals^{36,37}. It suggests that IL-1 dependent upregulatory mechanism of MMP-9 expression degrades the collagen fragments and extracellular matrix. The bone resorption by IL-1, IL-6, TNF- α and MMP-9 favours the expansion of periapical cyst ²⁷. The present study also represents positive correlation among IL-6, TNF-α and MMP-9 (IL-6 vs MMP-9, r=0.514* and TNF-α vs MMP-9, r= 0.768**). The results of present study support the view that lipopolysaccharide may play a prominent role in the pathogenesis of radicular cyst. It triggers the production of proinflammatory cytokines (IL-1, IL-6 and TNF- α) and MMP-9 from macrophages and other inflammatory cells at the site of inflammation and

stimulates the bone resorption by activation of osteoclastic activity (LPS vs IL-1, r= 0.661^{**} and IL-1 vs MMP-9, r= 0.715^{**}). Further investigation may explore the complex mechanism involved in the development of radicular cyst and periapical bone resorption.

CONCLUSION

Present study indicates the significant correlation of LPS with the pathogenesis of radicular cyst. Moreover, it indicates that due to increased bacterial toxins in the root canal system the expression of several interleukins and MMP-9 are also overexpressed that contribute the periapical bone resorption and hence take part in the pathogenesis of radicular cyst.

Acknowledgements: The authors are highly thankful to Prof. Dr. M. H. Qazi, Center for Research in Molecular Medicine (CRiMM) to provide the innovative and financial support for the project Conflict of Interest: Authors declare no conflict of interest.

Authors Contribution

AQ, AS, KA writing the article and reviewing the sources

NT, SA, NS Resourcing materials, practical performance.
 SZ, SW, AM Conceptualization, performing statistics, reviewing and editing the article.

REFERENCES

- Leonardo MR, Silva LAB, Leonardo RT. Tratamento de canal radicular em sessão única: crença vs. ciência. In: Feller C, Gorab R. Atualização na clínica odontológica. São Paulo: Artes Médicas. 2000; 3:27-57.
- Gomes BP, Endo MS, Martinho FC. Comparison of endotoxin levels found in primary and secondary endodontic infections. J Endod. 2012; 38(8):1082-6.
- Lin SK, Kok SH, Lin LD, Wang CC, Kuo MYP, Lin CT, Hsiao M, Hong CY. Nitric oxide promotes the progression of periapical lesion via inducing macrophage and osteoblast apoptosis. Oral Microbiol Immunol. 2007; 22(1): 24-29.
- Paula-Silva FW, Da Silva LA, Kapila YL. Matriz metalloproteinase expression in teeth with periapical periodontitis is differentially modulated by the modality of root canal treatment. J Endod. 2010; 36(2):231-7.
- Leonardo MR, Leonardo RT, Nelson-Filho P, Ferrari PHP. A endotoxina e sua importância na infecção endodôntica. Em: Ferrari PHP, Bombana AC. A Infecção endodôntica e sua resolução. São Paulo: Editora Santos. 2010; 47-61.
- Mohammadi Z. Endotoxin in endodontic infections: a review. J Calif Dent Assoc. 2011; 39(3):152-5.
- 7. Anas A, Pol TV, De-Vos AF. Role of CD14 in lung inflammation and infection. Critical Care. 2010; 14(2):209.
- Stefan A, Hienz, Sweta Paliwal, Saso Ivanovski. Mechanisms of Bone Resorption in Periodontitis Journal of Immunology. 2015; 1-10.
- Graves DT, Corchan D. The contribution of interleukin-1 and tumor necrosis factor in periodontal tissue destruction Periodontol. 2003; 74(3): 391-401.
- Teng Y, Nguyenn H, Gao X. Functional T-cell immunity and osteoprotegerin ligand control alveolar bone distruction in periodontal infection. J Clin Invest Dis. 2000; 106(6):59-67.
- Leonardo MR, Silva RAB, Assed S, Nelson-Filho P. Importance of bacterial endotoxin (LPS) in Endodontics. J Appl Oral Sci. 2004; 12(2):93-8.
- Poltorak A, He X, Smirnova I. Defective LPS signaling in C3H /HeJ and C57BL /10ScCr mice: mutations in the TIr4 gene. Science. 1998; 282(5396):2085-8.
- Baker P, Dixon M, Evans R, Dufour L, Johnson E, Roopenian D. CD4 (+) T cells and proinflammatory cytokines gamma interferon and interleukin-6 contribute to alveolar bone loss in mice. Infect Immun. 1999; 67(6):2804-9.
- Triantafilou M, Triantafilou K. Lipopolysaccharide recognition: CD14, TLRs and the LPS-activation cluster. Trends Immunol. 2002; 23(6):301-4.
- Muroi M, Ohnishi T, Tanamoto K. Regions of the mouse CD14 molecule required for toll-like receptor 2- and 4-mediated activation of NF-kappa B. J Biol Chem. 2002; 277(44):42372-9.
- Miyake K, Nagai Y, Akashi S, Nagafuku M, Ogata M, Kosugi A. Essential role of MD-2 in B-cell responses to lipopolysaccharide and Toll-like receptor 4 distribution. J Endotoxin Res. 2002; 8(6):449-52.
- Gusman H, Santana RB, M Zehnder. Matrix metalloproteinase levels and gelatinolytic activity in clinically healthy and inflamed human dental pulps. Eur J Oral Sci. 2002; 110(5):353-7.

- Ebersole JL, Dawson DR, Morford LA, Peyyala R, Miller CS, Gonzalez OA. "Periodontal disease immunology: "double indemnity" in protecting the host." Periodontology. 2013: 62(1):163-202.
- protecting the host," Periodontology. 2013; 62(1):163-202.
 Han X, Kawai T, Taubman MA. "Interference with immune-cell-mediated bone resorption in periodontal disease," Periodontology. 2007; 45(1):76-94.
- Jin Q, Cirelli JA, Park CH. RANKL inhibition through osteoprotegerin blocks bone loss in experimental periodontitis. Journal of Periodontology. 2007; 78(7):1300–1308.
- Prikk K, Maisi P, Pirila E, Sepper R, Salo T, Wahlgren J, Sorsa T. In vivo collagenase-2 (MMP-8) expression by human bronchial epithelial cells and monocytes/ macrophages in bronchiectasis. J Pathol. 2001; 194(2):232-238.
- Takeichi O, Saito I, Okamoto Y, Tsurumachi T, Saito T. Cytokine regulation on the synthesis of nitric oxide in vivo by chronically infected human polymorphonuclear leucocytes. Immunology. 1998; 93(2):275-80.
- Nagai Y, Akashi S, Nagafuku M. Essential role of MD-2 in LPS responsiveness and TLR4 distribution. Nat Immunol. 2002; 8(6)3:667.
- 24. Martinho FC, Chiesa WM, Zaia AA, Ferraz CC, Almeida JF, Souza-Filho FJ. Comparison of endotoxin levels in previous studies on primary endodontic infections. J Endod. 2011; 37(2):163-7.
- Underhill DM, Ozinsky A. Toll-like receptors: key mediators of microbe detection. Curr Opin Immunol. 2002; 14(1):103-10.
- Campos K, Gomes CC, Farias LC, Silva RM, Letra A, Gomez RS. DNA methylation of MMP9 is associated with high levels of MMP-9 messenger RNA in periapical inflammatory lesions. Journal of endodontics. 2016; 42(1):127-30.
- Corotti MV, Zambuzzi WF, Paiva KB, Menezes R, Pinto LC, Lara VS, Granjeiro JM. Immunolocalization of matrix metalloproteinases-2 and-9 during apical periodontitis development. archives of oral biology. 2009;54(8):764-71.
- Martinho FC, Chiesa WM, Leite FR, Cirelli JA, Gomes BP. Correlation between clinical/radiographic features and inflammatory cytokine networks produced by macrophages stimulated with endodontic content. Journal of endodontics. 2012. Jun 30;38(6):740-5.
- de Andrade SP, de Aquino AR, Oliveira BA, de Almeida FR, Galvao HC, de Souza LB. Immunohistochemical expression of nuclear factor kB, matrix metalloproteinase 9, and endoglin (CD105) in odontogenic keratocysts, dentigerous cysts, and radicular cysts. Oral surgery, oral medicine, oral pathology, oral radiology, and endodontics. 2011 ;112(4):476-83.
- Ragab AA, Nalepka JL, Bi Y, Greenfield EM. Cytokines synergistically induce osteoclast differentiation: support by immortalized or normal calvarial cells. American Journal of Physiology and Cell Physiology. 2002. 283; 679-87.
- Wan C, Yuan G, Yang J, Sun Q, Zhang L, Zhang J, Zhang L, Chen Z. MMP9 deficiency increased the size of experimentally induced apical periodontitis. Journal of endodontics. 2014;40(5):658-64.
- de Moraes M, de Lucena HF, de Azevedo PR, Queiroz LM, Costa AD. Comparative immunohistochemical expression of RANK, RANKL and OPG in radicular and dentigerous cysts. Archives of oral biology. 2011;56(11):1256-63.
- Hong CY, Lin SK, Kok SH, Cheng SJ, Lee MS, Wang TM, Chen CS, Lin LD, Wang JS. The role of lipopolysaccharide in infectious bone resorption of periapical lesion. Journal of oral pathology & medicine. 2004;33(3):162-9.
- Pattamapun K, Handagoon S, Sastraruji T, Gutmann JL, Pavasant P, Krisanaprakornkit S. Decreased Levels of Matrix Metalloproteinase-2 in Root-Canal Exudates During Root Canal Treatment. Archives of Oral Biology. 2017.
- Jain A, Bahuguna R. Role of matrix metalloproteinases in dental caries, pulp and periapical inflammation: an overview. Journal of oral biology and craniofacial research. 2015;5(3):212-8.
- Ahmed GM, El-Baz AA, Hashem AA, Shalaan AK. Expression levels of matrix metalloproteinase-9 and gram-negative bacteria in symptomatic and asymptomatic periapical lesions. Journal of endodontics. 2013;39(4):444-8.
- Kubota Y, Ninomiya T, Oka S, Takenoshita Y, Shirasuna K. Interleukin-1α-dependent regulation of matrix metalloproteinase-9 (MMP-9) secretion and activation in the epithelial cells of odontogenic jaw cysts. Journal of dental research. 2000;79(6):1423-30.
- Xie H, Chen P, Huang H W, Liu L P, Zhao F, Ning Z and Hong L. Reactive oxygen species downregulate ARID1A expression via its promoter methylation during the pathogenesis of endometriosis. Eur Rev Med Pharmacol Sci. 2007; 21(20), 4509